# **ORIGINAL ARTICLES**

## STUDIES ON THE OCCURRENCE OF GRAM-NEGATIVE BACTERIA IN TICKS: IXODES RICINUS AS A POTENTIAL VECTOR OF PASTEURELLA

Nimfa Maria Stojek, Jacek Dutkiewicz

Department of Occupational Biohazards, Institute of Agricultural Medicine, Lublin, Poland

Stojek NM, Dutkiewicz J: Studies on the occurrence of Gram-negative bacteria in ticks: *Ixodes ricinus* as a potential vector of *Pasteurella*. *Ann Agric Environ Med* 2004, **11**, 319–322.

Abstract: A total of 372 Ixodes ricinus ticks (101 females, 122 males, and 149 nymphs) collected by flagging in 6 mixed woodlands of eastern Poland were examined by culture for the presence of internal Gram-negative bacteria other than Borrelia burgdorferi. Adult ticks were examined in pools of 2 specimens each and nymphs were examined in pools of 3-5 specimens each. Ticks were disinfected in 70% ethanol and homogenized in 0.85% NaCl. The diluted homogenate was inoculated onto 3 kinds of agar media: buffered charcoal yeast extract (BCYE-a) for isolation of fastidious Gram-negative bacteria, eosin methylene blue agar (EMB) for isolation of enterobacteria, and tryptic soya agar for isolation of all other non-fastidious Gram-negative bacteria. The Gramnegative isolates were identified with the API Systems 20E and NE microtests. A total of 9 species of Gram-negative bacteria were identified, of which the commonest were strains determined as Pasteurella pneumotropica/haemolytica, which were isolated on BCYE-a agar from ticks collected in all 6 examined woodlands. The total number of these strains (49) exceeded the total number of all other strains of Gram-negative bacteria recovered from ticks (30). Of the total number of examined ticks, the minimum infection rate with Pasteurella pneumotropica/haemolytica was highest in females (18.8%), and slightly lower in males (12.3%) and nymphs (10%). Besides Pasteurella pneumotropica/haemolytica, the following species of Gram-negative bacteria were isolated from examined ticks: Pantoea agglomerans, Serratia marcescens, Serratia plymuthica on EMB agar and Aeromonas hydrophila, Burkholderia cepacia, Chromobacterium violaceum, Pseudomonas aeruginosa, and Stenotrophomonas maltophilia on tryptic soya agar. Minimal infection rates with these bacteria were low, ranging from 0.7-5.9%. Of the isolated bacteria, Chromobacterium violaceum, Pasteurella pneumotropica/haemolytica, Pseudomonas aeruginosa, and Serratia marcescens are potentially pathogenic for man and/or animals. In particular, the common occurrence of Pasteurella pneumotropica/haemolytica in Ixodes ricinus ticks poses a potential risk of pasteurellosis for humans and animals exposed to tick bites.

Address for correspondence: Dr Nimfa Maria Stojek, Department of Occupational Biohazards, Institute of Agricultural Medicine, Jaczewskiego 2, 20-090 Lublin, Poland. E-mail: nina@galen.imw.lublin.pl

Key words: ticks, *Ixodes ricinus*, transmission, Gram-negative bacteria, *Pasteurella*, *Chromobacterium*, *Pseudomonas*, *Serratia*.

#### **INTRODUCTION**

*Ixodes ricinus* L., inhabiting deciduous and mixed forests and surrounding areas, is the most common tick in Europe, including Poland [19]. It represents a significant

health hazard for humans and domestic animals as a vector of multiple pathogens, of which the most important are: *Flavivirus* causing tick-borne encephalitis, spirochete *Borrelia burgdorferi* causing Lyme disease, rickettsia *Anaplasma phagocytophilum* causing ehrlichiosis (ana-

Received: 10 September 2004

Accepted: 20 November 2004

plasmosis), Gram-negative bacterium *Francisella tularensis* causing tularemia, and protozoans *Babesia* spp. causing babesiosis in animals, rarely in humans [4, 8, 15, 16, 17, 20]. *Ixodes ricinus* has also been reported as participating in the transmission of many other pathogenic microorganisms, including: virus of Louping ill, virus of Crimean-Congo hemorrhagic fever, viruses of the groups Uukuniemi and Kemerovo, *Rickettsia slovaca, Coxiella burnetii, Salmonella enteritidis, Listeria monocytogenes, Erysipelothrix rhusiopathiae, Brucella melitensis, Theileria mutans, Theileria sergenti* [18]. Most probably this list is not yet complete, as recently we have detected DNA of *Toxoplasma gondii* in *Ixodes ricinus*, thus demonstrating the possibility of transmission of toxoplasmosis by these ticks [21].

The aim of the present work was to examine the role of *Ixodes ricinus* as a potential vector of Gram-negative bacteria pathogenic for humans and animals (except for *Borrelia burgdorferi* which will be a subject of a separate work) by recovery of these organisms from ticks collected in eastern Poland on 3 kinds of bacteriological media.

The preliminary results of this work have been presented elsewhere [22].

### MATERIALS AND METHODS

**Collection of ticks.** The infection of ticks with Gramnegative bacteria was studied in 6 localities (woodlands) of the Lublin province (eastern Poland) (Fig. 1). Ticks were collected by dragging a light woolen flag ( $60 \times 90$  cm) over the lower vegetation, mostly on forest paths, in glades, and at the edge of the woods.

All examined woodlands were mixed forests, composed mainly of English oak (*Quercus ruber*), Scots pine (*Pinus* sylvestris), European alder (*Alnus glutinosa*), European hornbeam (*Carpinus betulus*), white warty birch (*Betula* verrucosa), and European beech (*Fagus sylvatica*).

Ticks were collected during May 2003 and May 2004 in the following localities: 1) Dąbrowa – inside a suburban forest south of the city of Lublin, on forest paths; 2) Kozłówka – on the territory of a recreational woodland, in glades and on forest paths; 3) Rogóźno – on the territory of a lakeland northeast of Lublin, on the area surrounding the forest; 4) Krasne – on the territory of a lakeland in the vicinity of Rogóźno, in the area surrounding the forest; 5) Zwierzyniec – on the territory of the Roztoczański National Park, in glades and on forest paths; 6) Sobibór – in a woodland near the boundary of the country, on the area surrounding the forest. Collected ticks were placed in test tubes with a few blades of grass and preserved at room temperature until investigation.

**Isolation of bacteria from ticks.** Adult ticks (females and males) were examined for the presence of internal Gram-negative bacteria in pools of 2 specimens each and nymphs were examined in pools of 3-5 specimens each. Ticks were washed in saline (0.85% NaCl), submerged for 3 min in 70% ethanol for disinfection of external



Figure 1. Map showing area of study: Lublin province in eastern Poland.

surface, washed twice in saline, and finally triturated in 0.5 ml of saline with sterile glass homogenizer. The diluted homogenate was inoculated by spreading 0.1 ml aliquots on the agar surface with a sterile glass spatula, onto the following media:

• Buffered charcoal yeast extract (BCYE- $\alpha$ , Oxoid) for isolation of fastidious Gram-negative bacteria.

• Eosin methylene blue agar (EMB agar, Difco) for isolation of enterobacteria (*Enterobacteriaceae*).

• Tryptic soya agar (Difco) for isolation of all other non-fastidious Gram-negative bacteria.

The buffered charcoal yeast extract (BCYE- $\alpha$ ) agar is used mostly for isolation of *Legionella* [3, 23], but could be useful also for isolation of *Francisella tularensis* and related fastidious bacteria [23]. In this work, the charcoal yeast extract (CYE) agar base was used supplemented with the Growth Supplement SR 110 A (ACES buffer/potassium hydroxide, ferric pyrophosphate, Lcysteine HCl,  $\alpha$ -ketoglutarate) (Oxoid, England).

Inoculated BCYE-α, EMB and tryptic soya plates were incubated for 24-48 hrs at 37°C. Grown colonies were Gram-stained and Gram-negative strains were isolated on tryptic soya agar slopes.

The Gram-negative isolates were identified with the use of the API Systems 20E and NE microtests (bioMérieux, Marcy l'Etoile, France).

#### RESULTS

The results of the study are presented in Table 1. Altogether, 9 species were identified, of which 1 (*Pasteurella pneumotropica/haemolytica*) was isolated on BCYE-a agar, 3 (*Pantoea agglomerans, Serratia marcescens, Serratia plymuthica*) on EMB agar, and 5 (*Aeromonas hydrophila, Burkholderia cepacia, Chromobacterium violaceum, Pseudomonas aeruginosa, Stenotrophomonas maltophilia*) on tryptic soya agar (TSA). The species

Table 1. Gram-negative bacteria isolated from Ixodes ricinus ticks collected in various woodlands on the territory of Lublin province, eastern Poland.

Woodland		Dąb	orowa	Kozłówka			Rogóźno				K	rasne	Zwierzyniec			Sobibór				Total	
Stage	ŶŶ	33	NN	₽₽	33	NN	ŶŶ	33	NN	₽₽	33	NN	φç	33	NN	₽₽	33	NN	φç	33	NN
No. of examined ticks	10	5	13	8	10	15	16	22	10	11	11	5	36	34	73	20	40	33	101	122	149
BCYE-a agar																					
Pasteurella pneumotropica/ haemolytica complex	2 20	1 20	1 7.7	2 25	2 20	0	0	6 27.3	0	2 18.2	0	0	9 25	3 8.8	7 9.6	4 20	3 7.5	7 21.2	19 18.8	15 12.3	15 10
EMB agar																					
Pantoea agglomerans <sup>a</sup>	0	0	0	0	0	0	0	3 13.6	0	1 9.1	0	0	0	0	0	0	0	0	1 1	3 2.5	0
Serratia marcescens <sup>a,b</sup>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	6 30	4 10	0	6 5.9	4 3.3	0
Serratia plymuthica <sup>a</sup>	0	0	0	0	0	0	0	0	0	1 9.1	0	0	0	0	0	0	0	0	1 1	0	0
Tryptic soya agar																					
Aeromonas hydrophila	0	0	1 7.7	0	0	0	0	0	0	0	0	0	0	0	0	0	1 2.5	0	0	1 0.8	1 0.7
Burkholderia cepacia	0	0	1 7.7	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1 0.7
Chromobacterium violaceum <sup>b</sup>	0	0	0	0	0	0	0	0	0	0	0	0	1 2.8	4 11.8	0	0	0	0	1 1	4 3.3	0
Pseudomonas aeruginosa <sup>b</sup>	0	1 20	0	0	0	0	4 25	0	0	0	0	0	0	0	0	0	0	0	4 4	1 0.8	0
Stenotrophomonas maltophilia <sup>b</sup>	0	0	0	0	1 10	0	0	0	1 10	0	0	0	0	0	0	0	0	0	0	1 0.8	1 0.7

QQ = females,  $\partial \partial =$  males, NN = nymphs. In the upper part of each cell the minimal number of infected ticks is given, and in the lower part (italic) the calculated minimum infection rate (MIR, %). <sup>a</sup>Primary growth also on tryptic soya agar. <sup>b</sup>Primary growth also on BCYE- $\alpha$  agar.

isolated on EMB agar also showed an ability to grow on TSA, whereas some of the species isolated on TSA showed an ability to grow on BCYE- $\alpha$  agar (Tab. 1).

Strains identified as belonging to *Pasteurella pneumotropica/haemolytica* complex proved to be the most common Gram-negative bacteria occurring in *Ixodes ricinus* ticks. They were found in ticks from all the examined woodlands. *Pasteurella pneumotropica/haemolytica* was isolated more often from female and male ticks, infected in 5 out of 6 woodlands each, than from nymphs, infected in 3 out 6 woodlands. On average, the minimum infection rate (MIR) in females was 18.8%, in males 12.3%, and in nymphs 10%.

Other species of Gram-negative bacteria were isolated less frequently. Four of them (*Aeromonas hydrophila*, *Pantoea agglomerans*, *Pseudomonas aeruginosa*, *Stenotrophomonas maltophilia*), were found in ticks from 2 woodlands and 4 (*Burkholderia cepacia*, *Chromobacterium violaceum*, *Serratia marcescens*, *Serratia plymuthica*) in ticks from 1 woodland. Six isolations were made from males, 5 from females, and 3 from nymphs. Minimum infection rates were low, ranging from 0.7-5.9% (Tab. 1).

#### DISCUSSION

In the course of bacteriological examination of *Ixodes ricinus* ticks from eastern Poland, the common occurrence

of *Pasteurella* strains was found, not reported earlier from any of the hard ticks (Ixodidae) [4, 8, 15, 16, 17, 18, 20]. Until recently, only Martin & Schmidtmann [9] isolated a single strain determined as *Pasteurella* species B from *Ixodes scapularis* female, collected from deer in the USA. The strains determined in this study as *Pasteurella pneumotropica/haemolytica* were isolated from ticks collected in all examined localities, and their total number (49) exceeded the total number of all other strains of Gram-negative bacteria belonging to 8 species (30) recovered from these ticks.

The taxon *Pasteurella pneumotropica/haemolytica* used in the API identification system comprises 2 related species: *Pasteurella pneumotropica* and *Pasteurella* (*Mannheimia*) haemolytica. The inability of the determination of single *Pasteurella* species is a limitation of the API system, and in consequence, also of this study. Nevertheless, both *Pasteurella pneumotropica* and *Pasteurella haemolytica* are known as causative agents of disease in animals and occasionally in humans [1, 6, 7, 11, 24, 25]. Accordingly, even at the present identification level, the results of this study indicate a potential risk of pasteurellosis transmission by ticks.

*Pasteurella (Mannheimia) haemolytica* is an important cause of pneumonia in sheep and cattle [1, 6], while *Pasteurella pneumotropica* causes respiratory infection mostly in rodents [7]. Both species may cause infections

in humans [7, 11, 13, 14, 24], which are rare and usually do not involve the respiratory system [25]. Human infection with *Pasteurella pneumotropica* is often transmitted by bites of animals, mostly dogs and cats [7, 11, 14], similar to the well-known case of *Pasteurella multocida* [16]. The ease of *Pasteurella* transmission through injured skin suggests that such transmission could be possible also by tick bite. To check the potential significance of our findings for epidemiology of pasteurellosis in humans and animals, a continuation of this study is planned, including determination of *Pasteurella* isolates with the use of molecular methods.

Of the 9 species of Gram-negative bacteria isolated from *Ixodes ricinus* in the course of the present work, 4 species had been isolated from ixodid ticks by earlier authors: Burkholderia cepacia by Murrell et al. [12] from Ixodes holocyclus in Australia, Pantoea agglomerans (synonyms: Erwinia herbicola, Enterobacter agglomerans) by Murrell et al. [12] from Boophilus microplus in Australia, and by Dutkiewicz [5] from Dermacentor reticulatus in Poland, Pseudomonas aeruginosa by Amoo et al. [2] from Boophilus decloratus and Boophilus geigyi in Nigeria, and Stenotrophomonas maltophilia by Martin and Schmidtmann [9] from Ixodes scapularis in the USA and by Murrell et al. [12] from Ixodes holocyclus, Aponomma fimbriatum and Boophilus microplus in Australia. To the best of our knowledge, the isolation of Aeromonas hydrophila, Chromobacterium violaceum, Pasteurella pneumotropica/haemolytica, Serratia marcescens and Serratia plymuthica from ixodid ticks has not been reported previously.

Besides *Pasteurella pneumotropica/haemolytica*, at least 3 other species of Gram-negative bacteria isolated from *Ixodes ricinus* in the course of the present work may cause infectious disease in humans: *Chromobacterium violaceum*, *Pseudomonas aeruginosa*, and *Serratia marcescens*. *Chromobacterium violaceum* causes rare, but potentially fatal human infections in tropical and subtropical areas [10]. *Pseudomonas aeruginosa* and *Serratia marcescens* are known causative agents of purulent infections in man [25]. It is noteworthy that while *Pasteurella pneumotropica/haemolytica* was commonly distributed over the whole study area, *Chromobacterium violaceum* and *Serratia marcescens* showed a focal distribution and were isolated only from one locality each.

#### CONCLUSION

The strains of *Pasteurella*, identified as *Pasteurella pneumotropica/haemolytica*, proved to be the commonest Gram-negative bacteria occurring in *Ixodes ricinus* ticks collected in eastern Poland. This poses a potential risk of pasteurellosis for humans and animals exposed to tick bites in this area.

#### REFERENCES

1. Ackermann MR, Brogden KA: Response of the ruminant respiratory tract to *Mannheimia (Pasteurella) haemolytica. Microbes Infect* 2000, **2**, 1079-1088.

2. Amoo AO, Dipeolu OO, Akinboade AO, Adeyemi A: Bacterial isolation from and transmission by *Boophilus decoloratus* and *Boophilus geigyi. Folia Parasitol (Praha)* 1987, **34**, 69-74.

3. Atlas RM, Parks LC: *Handbook of Microbiological Media*. CRC Press, Boca Raton, FL 1993.

4. Balashov YS: Krovososushchye Kleshchi (Ixodoidea) - Perenoschiki Bolezney Cheloveka i Zhivotnyh (Blood-sucking Ticks (Ixodoidea) – Vectors of Human and Animal Diseases). Izdatelstvo Nauka, Leningrad 1967 (in Russian).

5. Dutkiewicz J: Studies on endotoxins of *Erwinia herbicola* and their biological activity. *Zbl Bakt Hyg I Abt Orig A* 1976, **236**, 487-508.

6. Ewers C, Lubke-Becker A, Wieler LH: *Mannheimia haemolytica* and the pathogenesis of enzootic bronchopneumonia. *Berl Munch Tierarztl Wochenschr* 2004, **117**, 97-115 (in German).

7. Frebourg NB, Berthelot G, Hocq R, Chibani A, Lemeland JF: Septicemia due to *Pasteurella pneumotropica*: 16S rRNA sequencing for diagnosis confirmation. *J Clin Microbiol* 2002, **40**, 687-689.

8. Kettle DS (Ed): *Medical and Veterinary Entomology*. 2nd ed. CAB International, Wallingford, UK 1995.

9. Martin PAW, Schmidtmann ET: Isolation of aerobic microbes from *Ixodes scapularis* (Acari: Ixodidae), the vector of Lyme disease in the eastern United States. *J Econ Entomol* 1998, **91**, 864-868.

10. Midani S, Rathore M: Chromobacterium violaceum infection. South Med J 1998, **91**, 464-466.

11. Minton EJ: Pasteurella pneumotropica: meningitis following a dog bite. Postgrad Med J 1990, 66, 125-126.

12. Murrell A, Dobson SJ, Yang X, Lacey E, Barker SC: A survey of bacterial diversity in ticks, lice and fleas from Australia. *Parasitol Res* 2003, **89**, 326-334.

13. Nimri LF, Rawashdeh M, Meqdam MM: Bacteremia in children: etiologic agents, focal sites, and risk factors. *J Trop Pediatr* 2001, **47**, 356-360.

14. Olson JR, Meadows TR: *Pasteurella pneumotropica* infection resulting from a cat bite. *Am J Clin Pathol* 1969, **51**, 709-710.

15. Petrishcheva PA: *Perenoschiki Vozbuditeley Prirodnoochagovyh Bolezney (Vectors of the Natural Foci Disease Agents)*. Gosudarstvennoe Izdatelstvo Medicinskoy Literatury, Moskva 1962 (in Russian).

16. Schwabe CW: Veterinary Medicine and Human Health. 3rd Ed. Williams & Wilkins, Baltimore 1984.

17. Service MW (Ed): *Encyclopedia of Arthropod-transmitted Infections of Man and Domesticated Animal.* CAB International, Wallingford, UK 2001.

18. Siuda K: Kleszcze (Acari: Ixodida) Polski. Część I. Zagadnienia Ogólne (Ticks (Acari: Ixodida) of Poland. Part I. General Problems). PWN, Warszawa 1991 (in Polish).

19. Siuda K: Kleszcze (Acari: Ixodida) Polski. Część II. Systematyka i Rozmieszczenie (Ticks (Acari: Ixodida) of Poland. Part II. Taxonomy and Distribution). Polskie Towarzystwo Parazytologiczne, Warszawa 1993 (in Polish).

20. Sonenshine DE: *Biology of Ticks*. Vol. 2. Oxford University Press, New York 1993.

21. Sroka J, Chmielewska-Badora J, Dutkiewicz J: *Ixodes ricinus* as a potential vector of *Toxoplasma gondii*. Ann Agric Environ Med 2003, **10**, 121-123.

22. Stojek NM, Dutkiewicz J, Zwoliński J: Izolacja Gram-ujemnych bakterii z kleszczy *Ixodes ricinus* z terenu południowo-wschodniej Polski (Isolation of Gram-negative bacteria from *Ixodes ricinus* ticks collected in southeastern Poland). **In:** Pet'ko B, Juriš P (Eds): *Klieštami Prenášané Infekčné Choroby a Iné Zoonózy: Zbornik z Konferencie so Zahraničnou Učasťou, Košice, Slovakia, 19-20 October 2001*, 90-93 (in Polish).

23. Stypułkowska-Misiurewicz H, Krogulska B, Pancer K, Matuszewska R: *Legionella* sp.: Laboratoryjne rozpoznawanie zakażeń u ludzi i wykrywanie w środowisku wodnym (*Legionella* sp.: Investigation of human infection and detection in environmental water). *Roczn PZH* 2001, **52**, 1-18 (in Polish).

24. Takeda S, Arashima Y, Kato K, Ogawa M, Kono K, Watanabe K, Saito T: A case of *Pasteurella haemolytica* sepsis in a patient with mitral valve disease who developed a splenic abscess. *Scand J Infect Dis* 2003, **35**, 764-765.

25. Zaremba ML, Borowski J: *Podstawy Mikrobiologii Lekarskiej* (*Fundamentals of Medical Microbiology*). PZWL, Warszawa 1994 (in Polish).